IRIDOIDS : THE STRUCTURE ELUCIDATION OF SPECIONIN BASED ON CHEMICAL EVIDENCE AND ¹H NMR ANALYSIS

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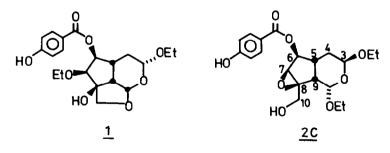
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Abstract : The iridoid specionin is an effective antifeedant against the Eastern spruce budworm. Previous synthetic work has shown that the proposed structure was incorrect. Presently the total synthesis of the revised structure $\underline{2C}$ is described. The structure elucidation, with special emphasis on the anomeric C-3 configuration, is based on chemical evidence and H MMR analysis.

INTRODUCTION

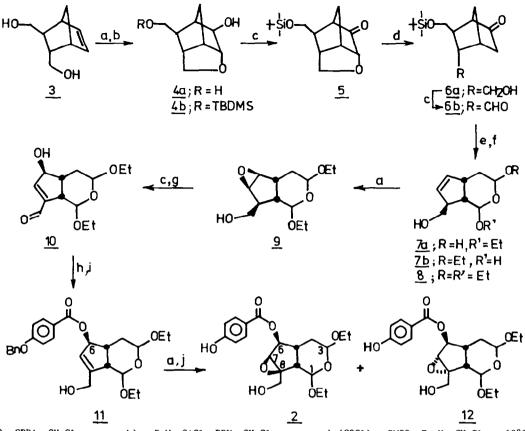
In 1983, the iridoid specionin has been isolated from the leaves of Catalpa speciosa Warder³. It has been shown to be an effective antifeedant against the Eastern spruce budworm which infests North American fir and spruce forests in May, inflecting huge damage to the lumber industry. Based on spectral data specionin has been given structure $\underline{1}^3$. In a previous paper⁴ we have reported the synthesis of substance $\underline{1}$ and found it not to be identical with the natural material. Comparison of the spectral data led us to propose structure $\underline{2C}$, with however unspecified configuration at C-3. In a preliminary note we have confirmed this structure (3-OEt unspecified) by total synthesis⁵. This synthesis afforded a mixture of isomers which could be separated by HPLC. One of the isomers was identical with an authentic sample of specionin. The relative stereochemistry shown for centers 1, 5, 6, 7 and 9 is based on NOE enhancements described by Chang and Nakanishi³; at that moment the configuration at C-3 could not be determined.



As already mentioned the synthetic material was produced together with isomers which were originally thought to be diastereoisomers at the anomeric positions C-1 and C-3. Reinvestigation of the synthesis has revealed that also isomers <u>12</u> (scheme 1) with a 7,8- α -oriented epoxide were formed. Although this is an unattractive aspect from the synthetic point of view it allowed complete configurational assignment of specionin as <u>2C</u>. Indeed only an extensive comparative study of the ¹H NMR spectra of the isomers <u>12</u>, coupled with chemical evidence allowed us to prove the β -orientation of the 3-ethoxysubstituent, while confirming the correctness for the other stereocenters.

Synthesis of specionin and of its isomers

The synthesis is based on a previously described strategy which allows a general entry into different subclasses of the iridoids via a Norrish I type fragmentation of suitably substituted norbornanone precursors 6 .



a) mCPBA, CH₂Cl₂, r.t.; b) t.BuMe₂SiCl, DBU, CH₂Cl₂, r.t.; c) (COCl₂, DMSO, Et₃N, CH₂Cl₂, -60°C; d) Al-Hg, EtOH, THF, r.t.; e) irradiation at 254 nm, EtOH; f) PTSA, EtOH, r.t.; g) DBU, CH₂Cl₂, r.t.; h) p.BnOC₆H₄COCl, Et₃N, CH₂Cl₂, r.t.; i) NaBH₄, EtOH, THF, 0°C; j) Pd-C, H₂, EtOH

SCHEME 1

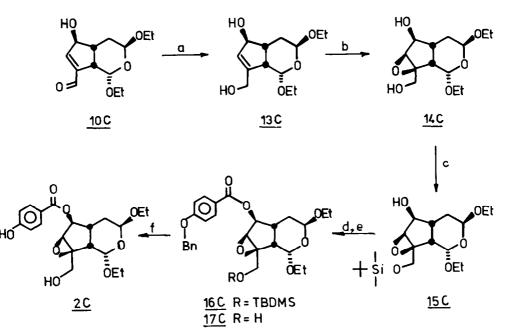
The starting material is the known norbornene $\underline{3}^{\prime}$ (Scheme 1). The keto-function in 6a is regioselectively introduced via a three step procedure with additional intermediate protection of m.Chloroperbenzoic acid induced cyclization of 3, via the the primary alcohol group in <u>4a</u>. elusive exo-epoxide, to 4a (96 %) followed by selective silyl ether 8 formation afforded 4b (81 %). Swern⁹ oxidation to 5 (77 %) and successive reductive cleavage of the lpha - ether bond led to <u>6a</u> (89 %). Swern oxidation of the hydroxyl function in <u>6a</u> yielded <u>6b</u> (89 %), the substrate for the Norrish I type fragmentation. Irradiation of a carefully degassed ethanolic solution of <u>6b</u> gave smooth fragmentation (TLC monitoring) of the norbornane framework. Most probably a diastereoisomeric mixture of 7a and 7b was formed as no aldehyde function could be detected. These intermediates could not be isolated as they are destroyed upon complete evaporation of the Therefore the ethanolic solution was treated with p.toluenesulfonic acid in order to solvent. perform the acetalization; concomitant silyl ether cleavage yielded directly a mixture of the four diastereoisomers 8 (64 %). The isomers could be separated by preparative HPLC; the ratio for <u>8C</u>, <u>8A</u>, <u>8D</u> and <u>8B</u> (for configurations see table 3) is 1:4:1:4 (order of elution). The relative configuration of the respective isomers $\underline{8}$ could not be determined unambiguously at this moment^{5,10}. Structural confirmation was only possible after transformation of each separate diastereoisomer into the final products 2 and 12 (vide infra).

The stage is now set for the functionalization of the cyclopentene ring. In order to prove the gross planar structure 2 proposed for specionin⁴, we decided to carry, in a first approach, the mixture of isomers 8 through the final steps. Epoxidation of 8 from the least hindered exo face gave 9 (94 %). Swern oxidation at -60°C (higher temperature as well as alternative pyridinium chlorochromate oxidation led to the formation of side products) followed by DBU mediated epoxide opening yielded the rather unstable aldehyde <u>10</u> (80 %). Formation of the 6-(pbenzyloxy)benzoate and reduction of the aldehyde function led to <u>11</u> (50 % overall). Finally

epoxidation of the isomers <u>11</u> (81 %) and deprotection of the phenolic function (100 %) gave a mixture of eight isomers (for the ratio see experimental part), consisting of two sets of compounds <u>2</u> and <u>12</u> with respectively a β - and α -oriented epoxide ring. This result shows that the last epoxidation step is not stereoselective; due to steric hindrance of the C-6 exo oriented ester group also epoxidation from the endo face has occurred.

HPLC separation allowed the isolation of synthetic (\pm) -specionin which was identical (¹H NMR spectroscopy, HPLC and GC retention times on co-injection) with a sample of the natural material kindly provided by Professor Nakanishi. This proves the gross planar structure for specionin; anticipating on the ¹H NMR analysis (vide infra) the natural product possesses the configuration <u>2C</u>.

We now faced the configurational assignment at the anomeric position C-3 and to a lesser extent at C-1. It should be stressed that, with the ¹H NMR spectrum of specionin only (or any other individual isomer <u>2</u> or <u>12</u>) in hand unambiguous structural determination of this conformationally flexible ring system is impossible. Evidently, the unselective epoxidation step is not attractive from the synthetic viewpoint, however it eventually allowed complete structural assignment of specionin (vide infra).



a) NaBH₄, EtOH, 0°C; b) mCPBA, CH₂Cl₂, r.t.; c) t.BuMe₂SiOTf, Et₃N, CH₂Cl₂, 0°C; d) p.BnOC₆H₄COC1, Et₃N, CH₂Cl₂, r.t.; e) HF, n.Bu₄N⁺F⁻, H₂O, THF, r.t.; f) Pd-C, H₂, EtOH SCHEME 2

Inspection of the ¹H NMR spectra of the isomers <u>2</u> and <u>12</u> indeed suggested that the configurations at the anomeric positions could be proven when also chemical evidence was taken into account. Therefore the four isomers <u>8</u> were taken separately through the reaction sequence shown in scheme 1. This allowed a classification of the isomers <u>2</u> and <u>12</u> in four pairs, each of them consisting of a β - and α -epoxide with identical configuration at the anomeric positions C-1 and C-3. For the sake of consistency the four isomers of intermediates <u>9</u>, <u>10</u> and <u>11</u> and of the final products <u>2</u> and <u>12</u> are also indicated as A, B, C and D (see table 3). The same results were found as starting from the isomeric mixture <u>8</u>, except for <u>8B</u> (1- and 3-OEt, β oriented) which upon epoxidation led to a mixture of <u>9B</u> and the isomeric 6,7- α -epoxide¹¹ (ratio 3:1).

Obviously, the formation of the isomers <u>12</u> is due to the steric hindrance exerted by the C-6 ester function in <u>11</u>. A stereoselective transformation of <u>8C</u> to specionin (<u>2C</u>) is possible via epoxidation of the allylic $alcohol^{12}$ <u>13C</u> (scheme 2). Reduction of the aldehyde <u>10C</u> gave the diol

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		<u>12A</u>	<u>12B</u>	9	<u>12C</u>		<u>12D</u>		<u>2A</u>		<u>2B</u>		50		<u>2</u> 0
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0 7 7	1.75		1.84	1.72	f1.77	2.04		2.26	1.8]	1.69	1.83	1.79	12.00	12.10	12.16
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Ś	2.48	2.36	2.86	I	2.69	2.63	2.39	2.08	2.44	2.51	2.51	2.49	2.45	2.67	2.35
9	5.06	5.02	5.07	5.12	5.07	5.07	5.07	5.01	5.59	5.89	5.26	5.26	5.37	5.61	5.29
7	3.71	3.46	3.68	3.49	3.56	3.38	3.57	3.31	3.79	3.66	3.72	3.51	3.77	3.64	3.83
6	2.60	2.60	2.70	2.71	2.69	2.38	2.68	2.37	2.47	2.47	2.51	2.44	2.81	2.84	2.69
10a	4.11	3.89	4.09	3.82	4.11	4.00	4.10	3.91	4.10	3.87	4.07	3.81	4.02	3.74	3.99
10b	3.77	3.63	I	3.55	3.78	3.83	3.85	3.77	3.63	3.58	3.65	3.56	3.74	3.40	I
OCH, CH,	3.97	3.79	3.98	I	4.10	3.85	4.08	3.87	4.05	3.82	4.06	3.84	3.87	3.79	I
ī	3.89	3.79	3.85	I	3.75	3.54	3.96	3.84	3.75	3.49	3.95	3.77	3.85	3.58	ı
F	3.63	3.30	3.58	I	3.59	3.19	3.59	3.31	3.56	3.14	3.59	3.30	3.51	3.22	I
-	3.54	3.26	3.48	ı	3.51	3.15	3.52	3.23	3.49	3.13	3.54	3.13	3.48	3.11	I
OCH, CH,	1.29	1.12	1.26	1.09	1.31	1.06	1.30	I	1.31	1.04	1.30	1.13	1.24	1.08	ı
4	1.23	1.03	1.19	1.08	1.21	1.02	1.23	ı	1.24	1.01	1.23	1.02	1.21	0.93	ı
arom	7.92	8.00	7.98	7.93	7.89	8.04	7.90	I	7.96	8.09	7.97	8.09	7.98	8.09	ł
	6.86	6,49	7.00	6.46	6.85	6.63	6.86	ı	6.86	6.50	6.87	6.48	6.87	6.43	1
^a For eac	h compou	nd the spe	For each compound the spectrum in $C_{\boldsymbol{6}}D_{\boldsymbol{6}}$ or		CDC1, sol	solution aff	ded a	complete se	set of coup	coupling constants by		simple first	t order me	order measurements,	s, thus
b Data n	g very ci	omplex sim red are in	voiding very complex simulations in č Data not massured are indicated by -	ňčase of ′–	degeneration of	ion of the	e spin system	tem.							
c Where	possible	, an assig	where possible, an assignment of the α - or	the a - or	<u>م</u>	i of the	H-4 proton is made	is made.							
	orts wer	e undertak	ten for as:	signment o	f the position of	tion of t	the UCH ₂ CH ₃ groups.	groups.							

Table 1 : ¹H NMR data of the isomers <u>2</u> and <u>12</u> (in CDC1, and C₆D₆ solution^a, TMS internal) : §-values (360 MHz)^b

E. VAN DER EYCKEN et al.

ł	<u>2D</u>	cDC1 ₃	3.1 3.7 3.3	9.4 6.7 9.2	1.5 1.5	, thus
						7 rements
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		CDC13	4.0 [3.0 13.9	5.1 7.2 8.4 8.2	12.5 12.5 12.5 12.5	8.8 st orde
		$c_6 D_6$	8.4 2.8 9 13.9	2.7 6.1 8.9	1.3 9.6 7.7.5	8.8 Imple fir
	<u>2B</u>	cDC13	8.4 2.7 9.2 14.1	1.9 5.9 9.1 -	1.3 9.4 7 7 7	8.8 ants by si
	<u>2A</u>	c ₆ b ₆	8.6 1 4.5 14.5	1 6.5 8.9	1.2 13.3 9.2 7.7	8.8 .ng const
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	۵	c ₆ b ₆	4.9 9.7 3.7 12.5	13.3 5.5 8.3 8.3	< 1 12.5 9.4 7.4	8.8 lete set m. s made.
	<u>12D</u>	cDC13	4.3 - -	- - 8.1	< 1 12.2 9.4 7	3.7 8.8 8.8 8.8 8.8 8.8 8.8 8.8 8.8 8.8 8
	<u>12C</u>	c ₆ D ₆	4.0 4.4 1.2 13.4	13.2 7.1 <1 7.8	< 1 12.7 9.2 7	8.8 on afford of the s the H-4
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	<u>12A</u>	c ₆ D ₆	7.7 8.7 6.3 -	14.4 5.5 <1 10.0	<pre>^ 1 12.9 9.5 7</pre>	8.7 spectrum simulati e indicat
		cDC1 ₃	7.7 9.0 6.0 13.9	14.4 5.8 <1 10.1	< 1 13.1 9.6 7	8.8 pound the y complex asured are
			³ J(1,9) 3J(3,4α) ^c J(3,4β) ^c J(4α4β)	J(4 0,5) ^c 3J(4 8,5) ^c 3J(5,6) 3J(5,9)	² 1(6,7) ² 1(10a,10b) ² 1(<u>0CH</u> ₂ ,CH ₃) ³ 1(0CH ₂ ,CH ₃)	arom 8.8 8.7 9 $\frac{1}{6}$ For each compound the spectrum in $C_{0}D_{c}$ or bavoiding very complex simulations in case bata not measured are indicated by Where possible, an assignment of the α -

Table 2 : ¹H NMR data of the isomers $\overline{2}$ and $\underline{12}$ (in CDCl₃ and C_6D_6 solution^a, TMS internal) : J-values (360 MHz)^b

Iridoids

<u>13C</u> (60 %) which upon epoxidation provided a single product <u>14C</u> (51 %). Protection of the primary alcohol function in <u>14C</u> led to <u>15C</u> (67 %) next to the disilylated product (26 %)¹³. The alcohol <u>15C</u> was transformed into the 6-(p-benzyloxy)benzoate <u>17C</u> (60 %) via esterification to <u>16C</u> and silyl ether cleavage¹⁴. Finally hydrogenolysis of the benzyl ether led to specionin <u>2C</u> (100 %).

Comparative ¹H NMR study of specionin and its isomers

Assignment of the orientation of the epoxide ring was straightforward and is based on the value of the coupling constant (table 2 and scheme 3) between H-5 and H-6. The four isomers 12 display a small ${}^{3}J(5,6)$ value (<1 - 2.3 Hz) which indicates a tortional angle of ca 90°. Knowing the 6-oxy function is β oriented (chemical evidence), this angle is obtained when this substituent and an α -epoxide oxygen atom are at maximum distance (electrostatic repulsion). For the β -epoxides 2, with an equatorially oriented 6-oxy function, the large ${}^{3}J(5,6)$ values of 6.7-8.9 Hz are in agreement with the antiperiplanar positions of H-5 and H-6. It should be mentioned that all naturally occurring 7,8-epoxy-iridoids display a β -epoxide function; it is thus logical to find specionin amongst the isomers 2.

<u>Table 3</u> : ¹H NMR 3 J values of the intermediates (CDCl₃; 200 MHz)^a

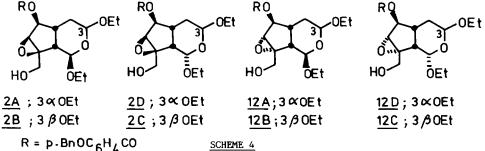
		Hax OE1 0	He Ha OEt	He Ha	Et			HO	H6 C1 C4
A	<u>B</u>		<u>C</u>	<u>D</u>			R = p.BnOC	6H4C0	
Product 8	<u>A</u>	<u>B</u>	<u>C</u>	<u>D</u>		<u>S</u>	<u>CHEME 3</u>		
(1,9) (3,4a) (3,4B) (4a,5) (4B,5)	7.5 [5.5 9 [5.5 -	5 [7.8 3.8 [6.5 5	3.3 [4.5 [4.3 [6.3 8	4 [2.5 9.5 [6 -		Ц		ŀ	1 5
Product 9						/ EtO		/	
(1,9) (3,4α) (3,4β) (4α,5) (4β,5)	3 9.3 3.8 13 5.5	2.5 [5.5 [4 [6.5 [8	3 4 1.5 12.5 5.8	3 [2.3 9.5 [-		10	High High High	<u>ــــ</u> ٥	EtO H4 p
Product 10						<u>12B</u>	<u>12C</u>	<u>12A</u>	<u>12D</u>
(1,9) (3,4α) (3,4β) (4α,5) (4β,5)	5 [4 [4 [6.5 [5.8	2 5 9 3 5	4 [4.8 3.3 [7.5 7	5 9.5 5 13 6.5	³ J(4α,5) 3J(4β,5) 3J(3,4α) 3J(3,4α) J(3,4β)	12.3 6.1 3.8 2.8	13.2 7.1 4.4 1.2	14.4 5.5 8.7 6.3	13.3 5.5 9.7 3.7
Product 11							SCHE	<u>ME 5</u>	
(1,9) (3,4a) (3,4B) (4a,5) (4B,5)	8 [5.5 7 [5.5 [10.5	6.5 [5.3 [- [-	4 6 6.5 6	5 9.5 4 13 5.8					

^a Assignment of the H-4 α and H-4 β was only possible for compounds <u>9A</u>, <u>9C</u>, <u>10D</u> and <u>11D</u>.

Considering the 3 J(1,9) values (table 2) the isomers <u>2</u> and <u>12</u> can be divided into two groups; <u>A</u> and <u>B</u> with large values (7.3-8.6 Hz) and <u>C</u> and <u>D</u> with smaller values (3.1-4.9 Hz). The large coupling constant is only plausible when H-1 and H-9 take an antiperiplanar position, thus

5390

with a 1 β -ethoxy substituent (isomers <u>2A</u>, <u>2B</u>, <u>12A</u> and <u>12B</u>). An analogous situation is observed for 2-deoxy-D-aldopentopyranoses or aldohexopyranoses¹⁵; for e,e or e,a dispositions the ³J(1,2) values vary between 2 and 4 Hz while for an a,a situation values of ca 9 Hz are found. On the other hand considering a 1^{α} -ethoxy group, for the isomers displaying the large J values, would necessitate the occurrence of a more energy demanding conformation with H-1 and H-9 close to an eclipsed position. Additional evidence is found in the ¹H NMR spectra (table 3) of the precursors <u>8</u> to <u>11</u>. The isomers <u>C</u> and <u>D</u> invariably show small coupling constants between H-1 and H-9 therefore excluding their antiperiplanar orientation and indicating a 1_{α}-ethoxy group. The C-1 configuration is furthermore supported by (a) the observation³ of a nuclear Overhauser effect between H-1 and H-9 for the naturally occurring isomer <u>2C</u>; (b) an independant synthesis¹⁶ of isomers <u>2A</u> and <u>2B</u> carried out by Prof. D. Curran; the β -orientation of the C-1-ethoxy group being ascertained by the nature of the starting material.



With all other stereocenters ascertained (scheme 4) we can now determine the configuration at C-3. It is obvious that as long as the conformation of the heterocyclic ring is unknown the coupling constants between H-3 α and H-4 β , respectively H-4 cannot be interpreted. Upo:: comparing the ${}^{3}J(4\alpha,5)$ and ${}^{3}J(4\beta,5)$ values observed for respectively the isomers 2 and 12 a neat distinction can be made (Table 2). The α -epoxides 12 display one large value, which indicates an antiperiplanar orientation between H-5 and one of the 4-protons. Thus, the heterocyclic part of the four isomers 12 seems to remain close to the extreme conformations I or II (scheme 5); in fact only the C-3, C-4, C-5 part has to be considered for the present discussion. This allows us to determine respectively H-4 α and H-4 β .

These informations combined with the ${}^{3}J(3,4\alpha)$ and ${}^{3}J(3,4\beta)$ values (scheme 5) can now be used for deducing the stereochemistry at C-3 for the isomers <u>12</u>. We have to refer again to the ${}^{3}J(1,2)$ values observed for the 2-deoxy-D-aldopentopyranoses and aldohexapyranoses (vide supra)¹⁵. The large ${}^{3}J(3,4\alpha)$ values found for <u>12A</u> (8.7 Hz) and for <u>12D</u> (9.7 Hz) indicate a trans diaxial coupling and hence an α -oriented ethoxy group. For the two other isomers <u>12B</u> and <u>12C</u> smaller coupling constants between H-4 α and H-3 are observed; therefore the 3-ethoxy group has a β -orientation.

Furthermore it is worth noting that a similar analysis of the coupling constants between H-3, H-4 and H-5 (table 2) observed for the β -epoxides <u>2A</u> and <u>2B</u> provides additional evidence for respectively a \Im - and 3β -ethoxy substituent; on the contrary no conclusion could be drawn from the ¹H NMR data of <u>2C</u> and <u>2D</u>.

However, as each isomer $\underline{12}$ is formed together with a β -epoxide partner $\underline{2}$ when starting from each individual isomer $\underline{8}$ (scheme 1), the structures $\underline{2A}, \underline{B}, \underline{C}$ and \underline{D} are also firmly established. In particular $\underline{2C}$ (identical to naturally occurring specionin) is obtained together with $\underline{12C}$, thus the 1 H NMR analysis and the chemical evidence unequivocally prove the structure of specionin.

Finally it should be mentioned that analysis of the ¹H NMR data of the intermediates <u>9A</u>, <u>9C</u>, <u>10D</u> and <u>11D</u> (table 3), as carried out for the compounds <u>12</u>, also allows unambiguous configurational assignment of the C-3 anomeric center. These results are in complete agreement with those obtained for the α -epoxides <u>12</u>. In retrospect, the present study leads to a reassignment of the configurations of the isomeric intermediates <u>8</u>¹⁰.

EXPERIMENTAL

IR spectra were recorded on a Beckmann IR 4230 spectrometer, mass spectra on a AEI MS-50 spectrometer. The H NMR spectra were recorded at 200 MHz (WH-Brucker) in CDCl₃ unless otherwise stated with TMS as internal standard. Chemical shifts (δ) are expressed in ppm. Rf values are quoted for Merck silica gel 60 GF₂₅₄ plates of thickness 0.25 mm. M.p.s. are uncorrected. The combined extracts were dried over MgSO₄ unless otherwise stated. The solvent was removed from the filtered solution on a rotary evaporator. Column chromatographic separations were performed on silica gel. HPLC separations were performed on RSiL 10 µm silica using a Waters M6000 A pump with a 50 x 0.8 cm column or a Knauer model 64 with a 25 x 2.2 cm column, both with PL-detection RI-detection.

<u>Diol 4a</u>

To a soln of diol $\underline{3}$ (57 g, 370 mmol) in dry CH₂Cl₂ (1500 ml) mCPBA (76.2 g, 370 mmol) was added in small portions at 0°C. After 20 h, at r.t., dimethylsulfide (6 ml) was added and the solvent was partially evaporated in vacuo. Then 10 % aq. NaOH was added until a pH of ca 10. A sat. aq. NaCl soln was added and the mixture was continuously extracted with Et₂O for several days. NaCl soln was added and the mixture was continuously extracted with Et₂O for several days. M.p. : 97°C; Rf (EtOAc) : 0.08; IR (KBr) : 3660-3040, 1480, 1420, 1365 cm⁻²; H NMR : 3.98 (dd, 1, J = 5 and 1.5 Hz), 3.81 (dd, 1, ²J = 8 Hz, J = 3.5 Hz), 3.71 (d, 1, ²J = 8 Hz), 3.56 (m, 3), 2.62 (m, 1), 2.12 (m, 1), 1.96 (m, 1), 1.93 (m, 1, ²J = 10 Hz), 1.65 (m, 1, ²J = 10), 1.65 (bs, 2), 1.42 (m, 1, J = 7.5 Hz); MS : m/z 170 (M+', 30), 139 (44), 99 (41), 95 (46), 70 (43), 69 (100); HRMS : calc. for $C_9H_{14}O_3$: 170.0923; found : 170.0943.

<u>t.Butyldimethylsilyl ether 4b</u> To a soln of <u>4a</u> (10 g, 59 mmol) in CH_2Cl_2 (500 ml) and DBU (10.6 ml, 71 mmol) was added a soln of t.BuMe_SiCl (10.5 g, 70 mmol) in CH_2Cl_2 (110 ml) at r.t.. The reaction mixture was stirred at r.t. for 10 h. After partially removal of the solvent the mixture was poured in water (200 ml). r.t. for 10 h. After partially removal of the solvent the mixture was poured in water (200 mi). The water layer was extracted with CH_2Cl_2 (4 x 50 ml). The collected organic fractions were washed with brine and dried. Solvent evaporation and column chromatography (hexane/EtOAc 3:1), yielded the product <u>4b</u> (13.5₁ g; 81 %). Rf (hexane/EtOAc 7:3) : 0.12; IR (neat) : 3600-3100, 1470, 1460, 1260, 1135 cm⁻¹; H NMR : 3.97 (dd, 1, J = 5 and 1.5 Hz), 3.79 (dd, 1, J = 4 Hz, J = 7.5 Hz), 3.69 (d, 1, J = 7.5 Hz), 3.51 (d, 2, J_2= 8 Hz), 3.50 (m, 1), 2.58 (m, 1), 2.10 (m, 1), 1.91-1.85 (m, 3, J = 6.5 and 4 Hz), 1.65 (m, 1, J = 11 Hz), 1.39 (m, 1, J = 8 Hz), 0.90 (s, 9), 0.05 (s, 6); MS : m/z : 227 (48), 105 (39), 99 (41), 75 (100); HRMS : calc. for $C_{11}H_{19}O_3Si$: 227 1103. found : 227.1091. 227.1103; found : 227.1091.

<u>Swern oxidation of 4b to the ketone 5</u> A soln of DMSO (17 ml, 240 mmol) in CH₂Cl₂ (80 ml) was added to a soln of oxalyl chloride (10.4 ml, 119 mmol) in CH₂Cl₂ (170 ml) at - 60° C. After 5 min a soln of the alcohol <u>4b</u> (11.1 g, 39 mmol) in CH₂Cl₂ (140 ml) was added dropwise and the mixture was stirred for 75 min at - 60° C. mmol) in CH₂Cl₂ (140 mI) was added dropwise and the mixture was stirred for 75 min at -60°C. Then Et₃N (71 ml, 509 mmol) was added and the mixture was warmed slowly to -10°C. The solution was poured into 10 % aq. HCl. After extraction with CH₂Cl₂, the combined organic layers were washed with brine, sat. aq. NaHCO₃ soln, and brine. Drying (Na₂SO₄), solvent evaporation and column chromatography (hexane/EtOAc 4:1) gave product $\frac{5}{2}$ (8.5 g; 77 %). M.p. : 50° C; Rf (hexane/EtOAc 7:3) : 0.27; IR (KBr) : 1760, 1470, 1460, 1255, 1120, 1100, 1070, 1045 cm⁻¹; H NMR : 4.04 (dd, 1, J = 4 Hz, J = 8 Hz), 3.89 (m, 1, J = 5.5 Hz), 3.84 (d, 1, J = 8), 3.58 (d, 2, J = 7.5 Hz), 3.00 (m, 1), 2.50 (m, 1), 2.33 (m, 1), 2.04 (m, 1, J = 11.5 Hz), 1.78 (m, 1, J = 7.5 Hz), 1.70 (m, 1, J = 11.5 Hz), 0.89 (s, 9), 0.06 (s, 6); MS : m/z 75 (28), 69 (100); HRMS : calc. for C₁₅H₂₆O₃Si : 282.1651; found : 282.1627.

<u>Reductive</u> <u>a-ether</u> <u>cleavage</u> <u>of</u> <u>ketone</u> <u>5</u> : <u>formation</u> <u>of</u> <u>6a</u>

To a susp of freshly prepared aluminum amalgam (from 30 g aluminum) in EtOH (17 ml) and THF (40 ml) was added a soln of 5 (7.5 g, 27 mmol) in EtOH (35 ml) and THF (80 ml) at r.t. The mixture was stirred for 2 h. The solids were centrifugated off and were washed several times with EtOAc. was stirred for 2 h. The solids were centrifugated off and were washed several times with EtOAc. After removal of the solvents of the first extract (THF/EtOH), the collected organic phases were washed with sat. aq. NH₂Cl and with brine. Work-up and column chromatography (hexane/EtOAc 6:4) yielded pure $\underline{6a}$ (6.7 g; 89 %). Rf (hexane/EtOAc 6:4) : 0.17; IR (neat) : 1750, 1260, 1120, 1090, 1065 cm⁻¹; H NMR : 3.75 (dd, 1, J = 5.5 Hz, J = 9.3 Hz), 3.67 (ddd, 1, J = 6 and 6 Hz), 2 J = 10 Hz), 3.50 (ddd, 1, J = 9 and 2.5 Hz, J = 10 Hz), 3.242 (dd, 1, J = 9.3 Hz, 2.10 (m, 1), 1.98 (m, 1, J = 18.3), 1.85 (m, 1), 2.13 (ddd, 1, J = 3.8 and 1 Hz, J = 10.8 Hz), 1.70 (m, 1), 0.91 (s, 9), 0.09 (s, 6); MS : m/z 227 (16, 209 (14), 107 (23), 105 (29), 75 (100); HRMS : calc. for $C_{11}H_{19}O_3Si$: 277.1118. 227.1103; found : 227.1118.

 $\frac{5-\text{Endo-formyl-6-exo-tert.butyldimethylsilyloxymethylbicyclo}{2.2.1} \frac{1}{100} \frac{1}{2.2.1} \frac{1}{2$

Norrish type I fragmentation of 6b and subsequent acetalization to 8

A soln of <u>6b</u> (200 mg, 0.71 mmol) in carefully degassed EtOH (100 ml) was irradiated for 4 h at r.t. (Ultraviolet Products PCQ-X1 photochemical reactor; 253.7 nm). Then a catalytic amount of PTSA was added and the mixture was stirred for 24 h. After adding sat. aq. NaHO₃ (30 ml) the EtOH was removed in vacuo at 25°C. The water phase was extracted with Et_2^0 (3 x). Work-up and column chromatography (hexane/EtOAc 6:4) afforded a mixture of the four isomers <u>8</u> (<u>A</u>, <u>B</u>, <u>C</u> and <u>D</u>,

column chromatography (hexane/EtOAc 6:4) afforded a mixture of the four isomers <u>B</u> (<u>A</u>, <u>B</u>, <u>C</u> and <u>D</u>, ratio 4:4:1:1) (110 mg; 64 %). The isomers were separated by HPLC using (hexane/EtOAc/acetone 8:1.5:0.5) leading to pure <u>C</u>, a mixture of <u>A</u> + <u>D</u> and pure <u>B</u> (order of elution). A further separation of <u>A</u> and <u>D</u> was performed with (hexane/EtOAc 8:2). <u>8A</u> : Rf (hexane/EtOAc 6:4) : 0.22; IR (neat) : 3600-3100, 1145, 1100, 1055, 1000, 970 cm⁻¹; ¹H NMR : 5.76 (ddd, 1, J = 5.5, 2.5 and 2.5 Hz), 5.57 (ddd, 1, J = 5.5, 1.5 and 1.5 Hz), 4.97 (dd, 1, J = 9 and 5.5 Hz), 4.84 (d, 1, J = 7.5 Hz), 3.92 (dq, 2, J = 9.5 and 7 Hz), 3.58 (dq, 1, J = 9.5 and 7 Hz), 3.56 (dq, 1, J = 9.5 and 7 Hz), 3.45 (dd, 1, J = 8.5 Hz, J = 10.5 Hz), 2.88 (m, 2), 2.12 (ddd, 1, J = 7.5, 7.5 and 9 Hz), 2.04 (ddd, 1, J = 14 Hz, J = 5.5 and 5.5 Hz), 1.30 (m, 1), 1.26 (t, 3, J = 7 Hz), 1.23 (t, 3, J = 7 Hz); MS : m/z 242 (M⁺⁺, 1), 241 (2), 197 (13), 96 (100); HRMS : calc. for C₁H₂₁O₄ : 241.1440; found : 241.1444. <u>8B</u> : Rf (hexane/EtOAc 6:4) : 0.19; H NMR : 5.72 (ddd, 1, J = 5.5, 1.8 and 1.8 Hz), 5.72 (ddd, 1, J = 5.5, 2 and 2 Hz), 4.75 (ddd, 1, J = 8, 3.8 and 0.8 Hz), 4.60 (d, 1, J = 5.Hz), 3.93 (dq, 1, J = 9.5 and 7 Hz), 3.90 (dq, 1, J = 9.3 and 7 Hz), 3.65 (dd, 1, J = 5.5 Hz, 'J = 10.5 Hz), 3.93 (dq, 1, J = 9.5 and 7 Hz), 1.73 (ddd, 1, J = 8, 3.48 (dq, 1, J = 9.3 and 7 Hz), 3.50 (dq, 1, J = 9.5 and 7 Hz), 3.23 (m, 1), 2.81 (m, 1), 2.25 (ddd, 1, J = 9.3 and 7 Hz), 2.10 (ddd, 1, J = 8 and 6.5 Hz, 'J = 13.8 Hz), 1.73 (ddd, 1, J = 5 and 3.8 Hz, 'J = 13.8 Hz), 1.23 (t, 3, J = 7 Hz).

3, J = 7 Hz).

3, J = 7 Hz). <u>8C</u>: Rf (hexane/EtOAc 6:4): 0.24; ¹H NMR: 5.79 (ddd, 1, J = 5.5, 2.5 and 2.5 Hz), 5.55 (m, 1, J = 5.5 Hz), 5.10 (d, 1, J = 3.3 Hz), 4.93 (dd, 1, J = 4 and 4 Hz), 4.00 (dq, 1, J = 9.5 and 7 Hz), 3.83 (dq, 1, J = 9.5 and 7 Hz), 3.59 (dq, 1, J = 9.5 and 7 Hz), 3.49 (dq, 1, J = 9.5 and 7 Hz), 3.16 (m, 2), 2.19 (ddd, 1, J = 8, 7and 3.3 Hz), 1.82 (ddd, 1, J = 6.3 and 4.5 Hz, J = 14 Hz). 1.52 (ddd, 1, J = 8 and 4.3 Hz), $J_J = 14$ Hz), 1.27 (t, 3, J = 7 Hz), 1.23 (t, 3, J = 7 Hz). <u>8D</u>: Rf (hexane/EtOAc 6:4): 0.22; H NMR: 5.85 (ddd, 1, J = 5.5, 2.5 and 2.5 Hz), 5.55 (m, 1, J = 5.5 Hz), 4.84 (d, 1, J = 4 Hz), 4.54 (dd, 1, J = 9.5 and 2.5 Hz), 4.07 (dq, 1, J = 9 and 7 Hz), 1.92 (ddd, 1, J = 6 and 2.5 Hz, J = 13 Hz). The isomers 8 gave almost identical IR MS and HPMS spectra

The isomers $\underline{8}$ gave almost identical IR, MS and HRMS spectra.

Epoxide 9

 $\frac{\text{Epoxide 9}}{\text{A soln of 8}} (39 \text{ mg, 0.16 mmol) and mCPBA (70 \text{ mg, 0.32 mmol) in CH}_{2}Cl_{2} (8 \text{ ml) was stirred for 5 h at r.t.. Then sat. aq. NaHCO₃ and excess dimethylsulfide was added and the mixture was stirred for 10 min. After extraction with Et_{0}, the organic layer was washed with sat. aq. NaHCO₃ and brine. Work-up and column chromatography (hexane/EtOAc 6:4) yielded 9 (39 mg; 94 %).$ 9A : Rf (hexane/EtOAc 6:4) : 0.07; IR (neat) : 3600-3100, 1150, 1100, 1050, 1015 cm²; ¹ H NMR : 4.90 (dd, 1, J = 9.3 and 3.5 Hz), 4.82 (d, 1, J = 3 Hz), 3.91 (dq, 1, J = 9.3 and 7 Hz), 3.84 (dq, 1, J = 9.3 and 7 Hz), 3.82 (d, 2, J = 6.3), 3.54 (dq, 1, J = 9.3 and 7 Hz), 3.51 (dd, 1, J = 2.8 and 1.5 Hz), 3.50 (dq, 1, J = 9.5, 1.5 and 6.3 Hz), 1.89 (ddd, 1, J = 5.5 and 3.5 Hz, J = 13 Hz), 1.65 (ddd, 1, J = 9.5, 7.5 and 3 Hz), 1.24 (t, 3, J = 7 Hz), 1.22 (t, 3, J = 7 Hz); MS : m/z 258 (M⁺, 1), 257 (1), 213 (5), 95 (14), 81 (35), 72 (100); HRMS : calc. for C₁₃H₂₁O₅ : 257.1389; found : 257.1401.9B : Rf (hexane/EtOAc 6:4) : 0.07; ¹ H NMR : 4.82 (dd, 1, J = 5.5 and 4 Hz), 4.68 (d, 1, J = 2.5

257.1389; found : 257.1401. <u>9B</u>: Rf (hexane/EtOAc 6:4) : 0.07; ¹H NMR : 4.82 (dd, 1, J = 5.5 and 4 Hz), 4.68 (d, 1, J = 2.5 Hz), 3.88 (dq, 1, J = 9.5 and 7 Hz), 3.87 (dq, 1, J = 9.25 and 7 Hz), 3.81 (d, 2, J = 6.3 Hz), 3.56 (dd, 1, J = 2.5 and 1.5 Hz), 3.50 (dq, 1, J = 9.5 and 7 Hz), 3.47 (dq, 1, J = 9.3 and 7 Hz), 3.34 (bd, 1, J = 2.5 Hz), 2.81 (ddd, 1, J = 8.3, 8 and 6.5 Hz), 2.16 (ddt, 1, J = 8.3, 1.5 and 6.3 Hz), 2.02 (ddd, 1, J = 6.5₂and 5.5 Hz, J = 13.8 Hz), 1.80 (ddd, 1, J = 8.3, 8.3 and 2.5 Hz), 1.50 (ddd, 1, J = 4 and 8 Hz, J = 113.8 Hz), 1.22 (t, 3, J = 7 Hz), 1.20 (t, 3, J = 7 Hz). <u>9C</u>: Rf (hexane/EtOAc 6:4) : 0.07; ¹H NMR : 5.05 (d, 1, J = 3 Hz), 4.98 (d, 1, J = 4 and₂1.5 Hz), 3.99 (dq, 1, J = 9 and 7 Hz), 3.78 (dq, 1, J = 9.5 and 7 Hz), 3.75 (dd, 1, J = 8 Hz, ²J = 10.5 Hz), 3.55 (dd, 1, J = 9 and 7 Hz), 3.52 (dq, 1, J = 9.5 and 7 Hz), 3.41 (bd, 1, J = 2.5 Hz), 3.29 (dd, 1, J = 2.5 Hz), 2.79 (ddd, 1, J = 6.3, 6.3 and 12.5 Hz), 2.36 (dddd, 1, J = 2.9.3, 8, 4 and 1.51 Hz), 1.78 (ddd, 1, J = 9.5, 6.5 and 3 Hz), 1.71 (ddd, 1, J = 5.8 and 1.5 Hz, J = 13.5 Hz), 1.31 (ddd, J = 12.5 and 4 Hz, J = 13.5 Hz), 1.26 (t, 3, J = 7 Hz), 1.24 (t, 3, J = 7 Hz). <u>9D</u>: Rf (hexane/EtOAc 6:4) : 0.07; ⁺H NMR : 4.73 (d, 1, J = 3 Hz), 4.54 (dd, 1, J = 9.5 and 2.3 Hz), 1.78 (ddd, 1, J = 9.5, 6.5 and 3 Hz), 1.71 (ddd, 1, J = 5.8 and 1.5 Hz, J = 13.5 Hz), 1.31 (ddd, J = 12.5 and 4 Hz, J = 13.5 Hz), 1.26 (t, 3, J = 7 Hz), 1.24 (t, 3, J = 7 Hz). <u>9D</u>: Rf (hexane/EtOAc 6:4) : 0.07; ⁺H NMR : 4.73 (d, 1, J = 3 Hz), 4.54 (dd, 1, J = 9.5 and 2.3 Hz). Hz).

The isomers 9 gave almost identical IR, MS and HRMS spectra.

<u>α - β-Unsaturated γ -hydroxy aldehyde 10</u> A soln of DMSO (13 μ1, 0.182 mmol) in CH₂Cl₂ (0.5 ml) was added to a soln of oxalylchloride (8 μl, 0.093 mmol) in CH₂Cl₂ (0.5 ml) at -60°C. The mixture was stirred for 2 min, then a soln of <u>9</u> (10 mg, 0.039 mmol) in CH₂Cl₂ (1 ml) was slowly added. After 25 min at -60°C Et₃N (64 μ1, 0.460 mmol) was added and stirring was continued for 45 min. Then water (1.5 ml) and Et₂O (1.5 ml) were added and after stirring for 10 min the product was extracted with Et₂O. The organic layer was washed with brine and dried. After solvent evaporation in vacuo at 25°C, the crude product was taken up in CH₂Cl₂ (3 ml) and DBU (1.7 μl, 0.116 mmol) was added. This mixture was stirred for 2 h at r.t.. Work-up and column chromatography (hexane/EtOAc 6:4 and afterwards EtOAc) vielded pure 10 (8 mo: 80 %). yielded pure <u>10</u> (8 mg; 80 %).

 $\begin{array}{l} \hline 10A_1 : \text{ Rf (hexane/EtOAc 3:7) : 0.23; UV (MeOH) : } \lambda_{\text{max}} = 240 \text{ nm; IR (neat) : } 3600-3100, 1680, 1625 \\ \hline 10A_1 : \text{ Rf (hexane/EtOAc 3:7) : 0.23; UV (MeOH) : } \lambda_{\text{max}} = 240 \text{ nm; IR (neat) : } 3600-3100, 1680, 1625 \\ \hline 10A_1 : \text{ H NMR : } 9.82 (s, 1), 6.84 (dd, 1, J = 1.5 and 1.5 Hz), 5.02 (m, 1), 5.00 (dd, 1, J = 4 and 4 Hz), 4.95 (d, 1, J = 5 Hz), 3.84 (dq, 1, J = 9.5 and 7 Hz); 3.80 (dq, 1, J = 9.5 and 7 Hz), 3.50 (dq, 1, J = 9.5 and 7 Hz), 3.48 (dq, 1, J = 9.5 and 7 Hz), 3.17 (dd, 1, J = 9.5 and 5 Hz + LR), 2. 51 (dddd, 1, J = 7, 6.8, 5.8 and 6.5 Hz), 2.01 (ddd, 1, J = 6.5 and 4 Hz, J = 14), 1.71 \\ \end{array}$

(ddd, 1, J = 5.8 and 4 Hz, 2 J = 14 Hz), 1.21 (t, 3, J = 7 Hz), 1.20 (t, 3, J = 7 Hz); MS : m/z 256 (M⁺, 1), 255 (1), 211 (7), 138 (24), 136 (24), 107 (31), 81 (24), 79 (28), 72 (100). 10B : Rf (hexane/EtOAc 3:7) : 0.18; H NMR : 9.80 (s, 1), 6.78 (dd, 1, J = 1.8 and 1.8 Hz), 4.99 (ddd, 1, J = 6,2.3 and 2.3 Hz), 4.83 (dd, 1, J = 5 and 9 Hz), 4.77 (d, 1, J = 2 Hz), 3.90 (dq, 1, J = 9 and 7 Hz), 3.86 (dq, 1, J = 9 and 7 Hz), 3.48 (dq, 1, J = 9.3 and 7 Hz), 3.30 (ddq, 1, J = 8.8, 3.5 aand 2 Hz), 2.52 (dddd, 1, J = 5.8, 6, 5 and 3 Hz), 2.30 (ddd, 1, J = 9 and 5 Hz, J = 14 Hz), 2.14 (ddd, 1, J = 5 and 3 Hz, J = 14 Hz), 2.00 (bs, 1), 1.22 (t, 3, J = 7 Hz), 1.21 (t, 3, J = 7 Hz). 10C : Rf (hexane/EtOAc 3:7) : 0.33; H NMR : 9.77 (s, 1), 6.77 (dd, 1, J = 2 and 1 Hz), 5.08 (d, 1, J = 9.5 and 7 Hz), 3.69 (dq, 1, J = 9.5 and 7 Hz), 3.49 (dq, 1, J = 7.2 and 2 Hz), 3.35 (dq, 1, J = 9.5 and 7 Hz), 2.46 (dddd, 1, J = 9.5 and 7 Hz), 2.16 (ddd, 1, J = 7 and 3 Hz, J = 13.5 Hz), 2.06 (ddd, 1, J = 7.5 and 4.8 Hz, J = 13.5 Hz), 1.21 (t, 3, J = 7 Hz), 1.05 (t, 3, J = 7 Hz). 10D : Rf (hexane/EtOAc 3:7) : 0.23; H NMR : 9.81 (s, 1), 6.74 (dd, 1, J = 2 and 2 Hz), 5.23 (d, 1, J = 9.5 and 7 Hz), 3.48 (dd, 1, J = 9.5 and 7 Hz), 3.49 (dq, 1, J = 9.5 and 7 Hz), 3.35 (dq, 1, J = 9.5 and 7 Hz), 2.46 (ddd, 1, J = 9.5 and 7 Hz), 1.21 (t, 3, J = 7 Hz), 1.05 (t, 3, J = 7 Hz). 10D : Rf (hexane/EtOAc 3:7) : 0.23; H NMR : 9.81 (s, 1), 6.74 (dd, 1, J = 2 and 2 Hz), 5.23 (d, 1, J = 9.5 and 7 Hz), 3.51 (dq, 1, J = 9.3 and 7 Hz), 3.34 (dq_21, J = 9.3 and 7 Hz), 3.76 (dq, 1, J = 9.5 and 7 Hz), 3.51 (dq, 1, J = 9.3 and 7 Hz), 3.34 (dq_21, J = 9.5 and 7 Hz), 2.34 (dddd, 1, J = 13.9, 6.5 and 3 Hz), 2.15 (ddd, 1, J = 6.5 and 5 Hz, J = 13 Hz), 2.00 (ddd, 1, J = 13 and 9.5 Hz, J = 13 Hz), 1.22 (t, 3, J = 7 Hz), 1.04 (t, 3, J = 7 Hz). The isomers 10 gave almost identical UV, IR and MS spectra.

<u>p. Benzyloxybenzoate 11</u> A soln of <u>10</u> (30 mg, 0.117 mmol), Et₃N (65 µl, 0.470 mmol) and p.BnOC₆H₄COC1 (58 mg, 0.234 mmol) in CH₂Cl₂ (8 ml) was stirred for 24 h at r.t.. Then an additional equal portion of Et₃N and of the acid chloride were added and stirring was continued for 24 h. EtOH (80 µl) was added and Δ from the acid chloride were added and stirring was continued for 24 h. EtOH (80 µl) was added and after 2 h the mixture was diluted with Et_20 and washed with sat. aq. NaHCO₂ and brine. After drying and solvent evaporation the residue was dissolved in EtOH (2 ml) and ³THF (2 ml). NaBH₄ (17.8 mg, 0.468 mmol) was added and the suspension was stirred for 30 min at 0°C. After addition of sat. aq. NH₂Cl and water (until the salts were dissolved) the mixture was extracted with Et₂O.

(17.8 mg, 0.468 mmol) was added and the suspension was stirred for 30 min at 0°C. After addition of sat. aq. NH₂Cl and water (until the salts were dissolved) the mixture was extracted with Et₂O. Work-up and HELC purification (hexane/EtOAC 7:3) afforded product <u>11</u> (27 mg; 50 %). <u>11A</u>: Rf (hexane/EtOAC 6:4) : 0.23; UV (MeOH) : $\lambda_{m} = 258$ nm; IR (neat) : 1705, 1680, 1605, 1580, 1510 cm⁻¹; H NMR : 7.96 (d, 2, J = 9 Hz + LR), 7.39 (m, 5), 7.34 (d, 2, J = 9 Hz + LR), 5.87 (m, 1), 5.60 (m, 1), 5.12 (s, 2), 5.00 (dd, 1, J = 7 and 5.5 Hz), 4.74 (d, 1, J = 8 Hz), 4.28 (bs, 2), 3.96 (dq, 1, J = 9.5 and 7 Hz), 3.85 (dq, 1, J = 9.5 and 7 Hz), 3.56 (dq, 1, J = 9.5 and 7 Hz), 3.07 (m, 1, J = 8 Hz), 2.57 (m, 1), 2.19 (ddd, 1, J = 5.5 and 5.5 Hz, J = 14 Hz), 1.21 (ddd, 1, J = 5.5 and 5.5 Hz, J = 14 Hz), 1.61 (ddd, 1, J = 10.5 and 7 Hz), 2.57 (m, 1), 2.19 (ddd, 1, J = 5.5 and 5.5 Hz, J = 14 Hz), 1.61 (ddd, 1, J = 10.5 and 7 Hz), 2.57 (m, 1), 2.19 (ddd, 1, J = 5.5 and 5.5 Hz, S. 85 (m, 2), 5.13 (s, 2), 4.88 (dd, 1, J = 5.3 and 5.3 Hz), 4.52 (d, 1, J = 6.5 Hz), 4.27 (bs, 2), 3.99 (dq, 1, J = 9.3 and 7 Hz), 3.93 (dq, 1, J = 9.3 and 7 Hz), 3.54 (dq, 1, J = 9.3 and 7 Hz), 3.53 (dq, 1, J = 9.3 and 7 Hz), 3.93 (dq, 1, J = 9.3 and 7 Hz), 3.54 (dq, 1, J = 9.3 and 7 Hz), 3.53 (dq, 1, J = 9.3 and 7 Hz), 2.06 (dd, 1, J = 6.5 Hz), 4.27 (bs, 2), 3.99 (dq, 1, J = 9.3 and 7 Hz), 2.88 (m, 2), 2.04 (m, 2, J = 5.5 Hz), 1.25 (t, 3, J = 7 Hz). 1.1C : Rf (hexane/EtOAc 6:4) : 0.24; H NMR : 7.97 (d, 2, J = 9 Hz + LR), 7.37 (m, 5), 6.97 (d, 2, J = 9 Hz + LR), 5.87 (m, 1), 5.74 (m, 1), 5.12 (s, 2), 5.06 (dd, 1, J = 6.5 and 4.8 Hz), 5.06 (d, 1, J = 6 and 4.8 Hz), 5.06 (d, 1, J = 4 Hz), 4.28 (bs, 2), 3.92 (dq, 1, J = 9.5 and 7 Hz), 3.85 (dq, 1, J = 14.3 Hz), 1.24 (t, 3, J = 7 Hz). 1.27 (t, 3, J = 7 Hz). 1.27 (t, 3, J = 7 Hz). 1.27 (t, 3, J = 7 Hz), 1.22 (t, 3, J = 7 Hz), 1.28 (dd, 1, J = 6 and 6 Hz, J = 14.3 Hz), 1.22 (t, 3, J = 7 Hz). 1.87 (ddd, 1, J = 5.4 and 7 Hz), 3.54 (dq, 1, J = 9.5 and 7 Hz), 3.54 (dq, 1, J = 9.5 and 7 Hz), 3.54 (dq, 1

Hz), 2.19 (ddd, 1, J = 6 and 4 Hz, 2 J = 13 1.22 (t, 3, J = 7 Hz), 1.21 (t, 3, J = 7 Hz).

The isomers 11 gave almost identical UV, IR, MS and HRMS spectra.

Isomers 2 and 12 A soln of 11 (9 mg, 0.019 mmol) and mCPBA (18.5 mg, 0.085 mmol) in CH_Cl_ (1.5 ml) was stirred for 24 h at r.t.. Then sat. aq. NaHCO₃ (2 ml) and dimethylsulfide (0.5 ml) were added and the mixture was stirred for 10 min. The system was extracted with Et₂0. The organic layer was washed with sat. aq. NaHCO₃ and brine. Work-up and column chromatographic purification (hexane/EtOAc 7:3) gave a mixture of α - and β -epoxides (7.5 mg; 81 %). This mixture was taken up in EtOH (0.750 ml) and added to a suspension of prehydrogenated Pd-C in EtOH (0.750 ml) in H₂ atmosphere. After stirring for 40 min at 1 atm solid NaHCO₃ was added and the mixture was filtered through a path of celite. Removal of EtOH at 25°C and column chromatography (hexane/EtOAc 6:4) yielded a mixture of 27 and 12 (6 mg; 90 %). Ratios in each pair of α - and β -epoxide : 2A/12A : 1, 2B/12B : 2, 2C/12C : 0.8, 2D/12D 0.1 (only traces of 2D could be detected). For the reaction leading to the mixture of all isomers a HPLC separation on a reversed phase 25 x 2.2 cm column RSiL-C₁₈-HL-D 10 um (MeOH/H₂0 55:45; 10 ml/min) (UV detection) gave pure samples of isomers 2 and 12 (A, B, C and D). 2C : Retention time : 5.8; UV (MeOH) : λ_{m} = 254 nm; IR (neat) : 1710, 1610, 1590, 1510, 1275 cm '; 'H NMR : see table 1 and 2; MS : m/Z 365 (1), 349 (1), 138 (13), 122 (12), 121 (100), 107 (12); HRMS : calc. for C₁₈H₂O₈ : 365.1236; found : 365.1214. Retention time of 2A, 4.6; 2B, 2.9; 2D, 4.8; 12A, 5.3; 12B, 5.3; 12C, 6.7 and 12D, 3.9. The isomers 2 and 12 gave almost identical UV, IR, MS and HRMS spectra.

Dio1 13C

To a soln of <u>10C</u> (20 mg, 0.078 mmol) in EtOH (1 ml) and THF (1 ml), NaBH, (1.5 mg, 0.039 mmol) was added and the mixture was stirred for 1 h at 0°C. Then sat. aq. NH,CI (1.5 ml) and Et₂O (2 ml) were added and after 10 min the mixture was extracted with Et₂O (8 x). The collected organic m1) were added and after 10 min the mixture was extracted with Et₂O (8 x). The collected organic layers were washed with brine and dried. Evaporation of the solvent and column chromatography (hexane/EtOAc 2:8) afforded $\underline{13C}$ (12 mg; 60 %). Rf (hexane/EtOAc 2:8): 0.11; IR (neat): 3600-3100, 1375, 1350, 1160, 1120, 1050 cm⁻¹; H NMR : 5.80 (dd, 1, J = 1.5 and 1.5 Hz), 5.02 (d, 1, J = 4 Hz), 4.97 (dd, 1, J = 5 and 5 Hz), 4.62 (bs, 1), 4.24 (bs, 2), 3.90 (dq, 1, J = 10.5 and 7 Hz), 3.85 (dq, 1, J = 10.5 and 7 Hz), 3.52 (dq, 1, J = 9.5 and 7 Hz), 3.52 (dq, 1, J = 9.5 and 7 Hz), 3.21 (m, 1), 2.45 (dddd, 1, J = 8.5, 5.5, 5.5 and 4.5 Hz), 2.06 (ddd, 1, J = 5.5 and 5 Hz, J = 14 Hz), 1.81 (ddd, 1, J = 5.5 and 5 Hz, J = 14 Hz), 1.23 (t, 6, J = 7 Hz); MS : m/z 72 (100) (100).

Epoxide 14C

A soln of $\underline{13C}$ (11 mg, 0.043 mmol) and mCPBA (18 mg, 0.085 mmol) in CH₂Cl₂ (2 ml) was stirred for l h at r.t.. Then sat. aq. NaHCO₃ and excess dimethylsulfide were added and the mixture was stirred for 10 min. After extraction with Et₂O, the organic layer was washed with sat. aq. NaHCO₃ and with brine. Drying, solvent evaporation and column chromatography (hexane/EtOAc 3:7) yielded $\underline{14C}_1$ (6 mg, 51 %). Rf (hexane/EtOAc 2:8) : 0.19; IR (neat) : 3600-3100, 1150, 1125, 1100, 1030 cm⁻¹; H NMR : 4.99 (d, 1, J = 4 Hz), 4.83 (dd, 1, J = 6.5 and 2.8 Hz), 4.14 (dd, 1, J = 7.5 and 1.5 Hz), 3.96_2 (d, 1, J = 12.5 Hz), 3.86 (dq, 1, J = 9 and 7 Hz), 3.81 (dq, 1, J = 9 and 7 Hz), 3.70 (d, 1, J = 12.5), 3.52 (d, 1, J = 1.5 Hz), 3.48 (dq, 1, J = 9 and 7 Hz), 3.46 (dq, 1, J = 7 Hz); MS : m/z 229 (26), 183 (39), 73 (100); HRMS : calc. for C₁₁H₁₇O₅ : 229.1075; found : 229.1121. A soln of 13C (11 mg, 0.043 mmol) and mCPBA (18 mg, 0.085 mmol) in CH₂Cl₂ (2 ml) was stirred for 229.1121.

<u>t.Butyl-dimethylsilyl</u> ether 15C To a soln of <u>14C</u> (21 mg, 0.077 mmol) and Et₃N (16.0 μ 1, 0.115 mmol) in CH₂Cl₂ (1.5 ml) was added dropwise, at $\overline{0^{\circ}C}$, t.BuMe₂SiOTf (19.3 μ 1, 0.084 mmol). After stirring for 30 min, brine was added dropwise, at 0°C, t.BuMe_SiOTf (19.3 µ1, 0.084 mmol). After stirring for 30 min, brine was added and the mixture was extracted with CH_Cl_. Drying, solvent evaporation and column chromatography (hexane/EtOAc 7:3) afforded 15C(20 mg, 67 %) next to 10 mg of the disilylated compound (26 \%). Rf (hexane/EtOAc 7:3) : 0.21; TR (neat) : 3600-3100, 1250, 1080, 1020, 990, cm⁻¹; H NMR : 5.05 (d, 1, J = 4), 4.82 (dd, 1, J = 6.5 and 2.5 Hz), 4.13 (m, 1), 4_212 (d, 1, 'J = 12), 3.86 (dq, 1, J = 9.3 and 7 Hz), 3.80 (dq, 1, J = 9.5 and 7 Hz), 3.54 (d, 1, J = 12 Hz), 3.49 (dq, 1, J = 9.3 and 7 Hz), 3.45 (dq, 1, J = 9.5 and 7 Hz), 3.36 (d, 1, J = 1.5 Hz), 2.74 (dd, 1, J = 7.3 and 4 Hz), 1.92 (3, m), 1.22 (t, 3, J = 7 Hz), 1.19 (t, 3, J = 7 Hz), 0.90 (s, 9), 0.08 (s, 3), 0.06 (s, 3); MS : m/z : 343 (3), 285 (3), 267 (4), 239 (14), 211 (23), 75 (100), 73 (49); HRMS : calc. for $C_{17}H_{31}O_5Si$: 343.1941; found : 343.1949.

Ester 17C

To a soln of <u>15C</u> (4 mg, 0.010 mmol) in CH_2Cl_2 (1.5 ml) was added Et_3N (5.8 µl, 0.041 mmol) and p.BnOC H₂COC1 (5.1 mg, 0.021 mmol). The mixture was stirred for 48 h and then a second portion of Et_3N and p.BnOC H₂COC1 was added. Stirring was continued for 24 h, EtOH (15 µl) was then added and after 2 h the mixture was diluted with Et_2O and washed with sat. aq. NHCO₃ and brine. added and after 2 h the mixture was diluted with Et_0 and washed with sat. aq. NaHCO₃ and brine. Drying, solvent evaporation and column chromatography (hexane/EtOAc 9:1) yielded crude <u>16C</u> (8 mg). Rf (hexane/EtOAc 8:2) : 0.40. To a soln of crude <u>16C</u> (8 mg) in THF (0.6 ml) was added 1.5 mol equiv. of fluoride as a 1:1 mixture of HF and $p.Bu_N^{\rm FF}$ [17 µ1, 0.020 mmol of a 1.2 M solution of F, prepared from 0.35 ml of 1.0 M n.Bu_N ^NF⁻ in THH and 0.25 ml of 1.4 M HF in THF (prepared from 0.5 ml of 47 % HF and 9.5 ml of THF). The reaction mixture was stirred at r.t. for 48 h, diluted with Et₂O and extracted with brine. Drying, solvent evaporation and column chromatography (hexane/EtOAc 7:3) afforded pure <u>17C</u> (3 mg, 60 %). Rf (hexane/EtOAc 6:4) : 0.14; UV (MeOH) : $\lambda_{\rm max} = 260$ nm; IR (neat) : 1710, 1600, 1580, 1505 cm⁻; H NMR : 8.03 (d, 2, J = 9 Hz + LR), 7.40 (m, 5), 7.00 (d, 2, J = 9 Hz + LR), 5.37 (dd, 1, J = 8, 3 and 1.5 Hz), 5.13 (s, 2), 5.06 (d, 1, J = 4 Hz), 4.88 (dd, 1, J = 6.5 and 3 Hz), 4.00 (d, 1, J = 12 Hz), 3.86₂(dq, 1, J = 9.5 and 7 Hz), 3.48 (dq, 1, J = 9.5 and 7 Hz), 2.80 (dd, 1, J = 8 and 4 Hz), 2.44 (dddd, 1, J = 9.5 and 7 Hz), 2.01 (ddd, 1, J = 5.5 and 3 Hz), J = 14 Hz), 1.86 (ddd, 1, J = 7 and 7 Hz, J = 14 Hz), 1.24 (t, 3, J = 7 Hz), 1.20 (t, 3, J = 7 Hz); MS : m/z 211 (12), 92 (11), 91 (100), 72 (14); HRMS : calc. for $C_{14}H_{11}O_2$: 211.0759; found : 211.0695.

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